

The Importance of Endothelial Reprogramming in Intracranial Atherosclerosis

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Abstract : Intracranial atherosclerosis (ICAS) develops through complex interactions between multiple cell types, primarily cerebrovascular endothelial cells and neuroglia. The pathophysiology of ICAS involves progressive vascular wall remodeling, characterized by the accumulation of inflammatory cells, activated mesenchymal cells, oxidized lipids, and altered extracellular matrix components within the arterial intima, resulting in atherosclerotic plaque formation. Endothelial dysfunction represents a key initiating mechanism in atherogenesis. Endothelial reprogramming encompasses phenotypic plasticity and functional alterations of endothelial cells, including endothelial-to-mesenchymal transition (EndMT), transcriptional factors, cell signaling, growth factors, miRNAs. In the central nervous system, endothelial cell phenotype is intrinsically linked to blood-brain barrier (BBB) integrity and function. Therapeutic modulation of endothelial reprogramming presents a potential non-pharmacological cellular intervention strategy for ICAS. This review examines the mechanistic roles of endothelial cells in intracranial atherosclerosis and synthesizes current evidence regarding endothelial reprogramming in ICAS pathogenesis.

Keywords : Intracranial atherosclerosis (ICAS), Endothelial-to-mesenchymal cell transition (EndMT), Blood brain barrier (BBB), Endothelial reprogramming

INTRODUCTION

Intracranial atherosclerosis (ICAS) is characterized by pathological arterial wall remodeling, involving progressive plaque accumulation and luminal stenosis within the cerebral vasculature, representing a primary etiology of ischemic stroke [1]. Atherosclerosis is reported to cause cerebral infarction related to ICAS in 20 to 40 out of 100,000 people worldwide [2]. ICAS accounts for 8~10% of all ischemic strokes, with a notably higher prevalence (30~50%)

in Asian populations [3]. Multiple risk factors contribute to ICAS pathogenesis, including advanced age, diabetes mellitus, hypertension, hypercholesterolemia, obesity, cardiovascular disease, and metabolic disorders [4-6]. The accumulation of sticky substances in the arteries, called plaque, narrows or occludes the blood vessels, preventing sufficient blood flow to the brain [7,8]. The gradual deposition of atheromatous plaques, comprising lipids, cholesterol, and inflammatory mediators, leads to hemodynamic alterations that significantly increase stroke risk [9,10].

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Endothelial cells, which compose the inner surface of blood vessels, play an important role in regulating the homeostasis of blood vessel walls [11]. Endothelial dysfunction represents an early pathophysiological marker in atherogenesis [12]. Within the central nervous system, endothelial cells are integral components of the blood-brain barrier (BBB), maintaining selective permeability and protecting neural tissue from potentially harmful blood-borne substances and immune cells [13]. Cerebrovascular endothelial dysfunction promotes atheromatous plaque formation and progression through multiple mechanisms, including inflammatory cascade activation, oxidative stress induction, and BBB disruption, ultimately contributing to stroke pathogenesis and other cerebrovascular disorders [14-16]. A key mechanism of endothelial dysfunction involves endothelial-to-mesenchymal transition (EndMT), wherein endothelial cells undergo phenotypic transformation, acquiring mesenchymal characteristics and functions [17,18]. EndMT facilitates plaque development and inflammatory responses, thereby accelerating atherogenesis [19]. In the cerebral vasculature, EndMT of brain endothelial cells (BECs) compromises BBB integrity, promoting immune cell infiltration and contributing to various neurological pathologies [20,21].

Endothelial reprogramming, including EndMT, plays a significant role in the pathogenesis of intracranial atherosclerosis. To mitigate its effects, it is crucial to restore endothelial cell function or regulate signaling pathways associated with EndMT. Endothelial reprogramming holds promise as a novel therapeutic target for the prevention, progression inhibition, and treatment of atherosclerosis. In this review, we highlight recent advances in the understanding of endothelial reprogramming as a strategy to address endothelial dysfunction in intracranial atherosclerosis.

ENDOTHELIAL DYSFUNCTION IN INTRACRANIAL ATHEROSCLEROSIS

The endothelium is crucial for maintaining the structural integrity of the vascular wall and plays a key role in the homeostasis of vascular functions, including the prevention of platelet aggregation, inhibition of thrombosis, and regulation of selective permeability [22-24]. Endothelial cells regulate blood flow by modulating vascular tone and selectively controlling the movement of ions, fluids, and other molecules through tight junctions [25]. Physiologically,

endothelial cells influence inflammation and the immune response by expressing cell adhesion molecules (CAMs), cytokines, and chemokines [26,27]. However, endothelial dysfunction leads to impaired endothelial cell function, which is associated with a reduction in nitric oxide availability, an imbalance between vasodilation and vasoconstriction, increased production of reactive oxygen species (ROS), and heightened oxidative stress [28-30]. The primary contributors to endothelial dysfunction include diabetes, obesity, hypertension, hyperglycemia, smoking, aging, and metabolic syndrome [31].

In the brain, intracranial endothelial cells, referred to as BECs, are integral components of the BBB. They play a critical role in protecting the brain from harmful external substances, regulating the entry and exit of molecules essential for brain metabolism, and maintaining homeostasis [32-34]. Dysfunction of intracranial endothelial cells leads to blood vessel narrowing, lipid modification, foam cell formation, an exacerbated inflammatory response, and platelet aggregation, which contribute to the formation of atherosclerotic plaques and are considered early indicators of intracranial atherosclerosis [35]. Lipid metabolism plays an essential role in the progression of atherosclerosis [36]. Abnormal lipid levels are observed in atherosclerosis patients, and low-density lipoprotein (LDL) transports lipids through the blood [37]. Under oxidative stress, ROS react with LDL and oxidize to form oxidized LDL (ox-LDL), which is ingested by macrophages to form foam cells [38]. Ox-LDL can promote atherosclerosis pathogenesis through endothelial cell damage, platelet activation, smooth muscle cell proliferation and migration [39-41]. Foam cells can contribute to atherosclerotic plaque formation and cause ischemic stroke. Within endothelial cells, nitric oxide (NO) is a reactive free radical that plays a vital role in maintaining vascular homeostasis by promoting vasodilation, inhibiting platelet aggregation, and exerting anti-inflammatory and antioxidant effects [42-44]. Endothelial dysfunction is associated with a reduction in NO production and bioavailability, leading to an imbalance in vascular homeostasis that promotes thrombosis and inflammation in the vascular wall, which can progress to vascular diseases such as atherosclerosis [45-47]. NO is synthesized by nitric oxide synthase (NOS) enzymes, which exist in three isoforms: endothelial nitric oxide synthase (eNOS), neuronal nitric oxide synthase (nNOS), and inducible nitric oxide synthase (iNOS), each with distinct functions and properties [48]. eNOS plays a crucial role

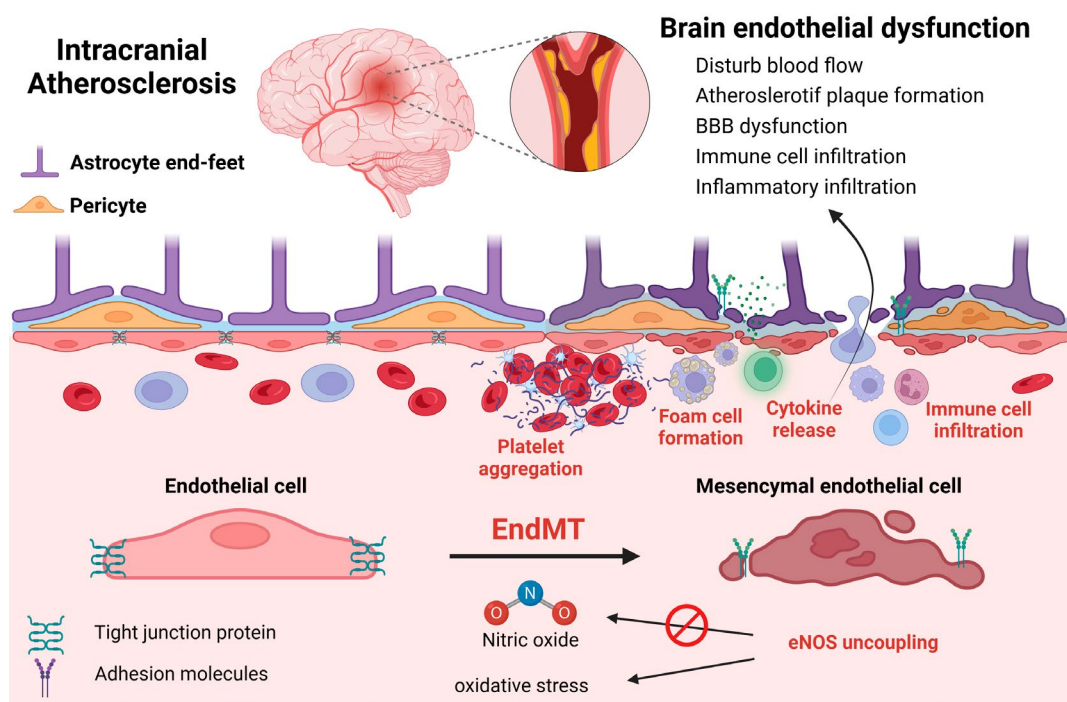


Fig. 1. Endothelial dysfunction in intracranial atherosclerosis. Figure showing the EndMT, the process by which endothelial cells transform into mesenchymal cells. EndMT results in decreased nitric oxide synthesis and increased oxidative stress due to eNOS uncoupling. Endothelial dysfunction leads to disturbance of blood flow, formation of atherosclerotic plaque, BBB dysfunction, immune cell infiltration and inflammatory cytokines and chemokines release.

in maintaining vascular homeostasis, and its deficiency is known to contribute to stroke development by impairing cerebral blood flow, promoting thrombosis, and inducing inflammation [49,50]. eNOS knockout (eNOS $-/-$) mice exhibit more severe neurological damage following stroke compared to control mice [51]. In the ischemic brain, progenitor cell proliferation and migration in the subventricular zone were reduced, and endothelial cell proliferation and vascular density were reduced at the ischemic border [51]. In eNOS $+/-$ mice, which exhibit partial eNOS deficiency, early thrombotic cerebral infarctions were observed, along with vascular occlusion in the temporoparietal and retrosplenial granular cortex, as well as the hippocampal regions [52]. Aged eNOS $+/-$ mice displayed BBB disruption, amyloid angiopathy, and cognitive decline [52]. eNOS $-/-$ mice subjected to bilateral common carotid artery stenosis (BCAS) exhibited reduced cerebral blood flow, glial activation, heightened inflammatory responses, BBB disruption, brain damage, and impaired cognitive function compared to control mice [53]. eNOS uncoupling occurs when electrons leak from the electron transport

chain and are transferred to oxygen molecules, generating superoxide (O_2^-) instead of NO [54]. Oxidative depletion of the eNOS cofactor tetrahydrobiopterin (BH4) and the depletion of the substrate L-arginine suggest that eNOS uncoupling induces oxidative stress in the brain, contributing to neurological damage [28,54]. These findings highlight NO as a crucial signaling molecule involved in various cerebrovascular diseases by regulating endothelial cell function. Modulating NO activity may offer an effective therapeutic strategy to protect against intracranial atherosclerosis by suppressing inflammatory responses, platelet aggregation, thrombosis, and oxidative damage, while preserving cerebral blood flow (Fig. 1).

CHANGED ENDOTHELIAL CELL IDENTITY IN INTRACRANIAL ATHEROSCLEROSIS

EndMT is a biological process in which endothelial cells acquire the functional and morphological characteristics of

mesenchymal cells, such as enhanced motility and contractility. Recent studies have linked EndMT to dysfunction of BECs [20,55]. In the brain, BEC dysfunction is associated with various neurological disorders, including multiple sclerosis, autoimmune encephalitis, and cerebral cavernous malformations, which result from BBB breakdown [56,57]. Intracranial atherosclerosis damages BECs, promotes plaque formation in blood vessels, and exacerbates BBB disruption, ischemic stroke, vascular dementia, and cognitive impairment [58-60]. During EndMT, BECs downregulate the expression of proteins that are critical for the structure and function of tight junctions, including claudin-5, claudin-11, claudin-25, occludin, zonula occludens-1 (ZO-1), ZO-2, junctional adhesion molecules (JAM-A, JAM-B, JAM-C), and adherens junction proteins such as α -catenin, β -catenin, platelet endothelial cell adhesion molecule-1 (PECAM-1), and vascular endothelial cadherin (VE-cadherin), thereby impairing the maintenance of barrier integrity [61,62]. Additionally, EndMT promotes BBB permeability by up-regulating the expression of adhesion molecules and matrix metalloproteinases, including vascular cell adhesion molecule-1 (VCAM-1), matrix metalloproteinase-3 (MMP3), MMP9, intracellular adhesion molecule-1 (ICAM-1), and monocyte chemoattractant protein-1 (MCP-1) [63,64].

EndMT induced by inflammatory responses is regulated by several signaling pathways, including tumor necrosis factor- α (TNF- α), transforming growth factor- β (TGF- β), WNT, vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF), NOTCH, and BMP2-JNK pathways [65-67]. TGF- β , a well-known inflammatory cytokine, is a key factor in inducing EndMT, which plays a critical role in acute vascular diseases and fibrosis [68]. In endothelial cells, TGF- β activation is mediated through type I receptors, specifically activin receptor-like kinases 1 and 5 (ALK1 and ALK5) [69]. Signal transduction from the TGF- β receptor complex to the nucleus is facilitated by Smad transcriptional regulators [70]. Smad2/3 are phosphorylated by ALK5, form a complex with Smad4, translocate to the nucleus, and regulate gene expression to promote EndMT [71]. Furthermore, inhibition of signal transducer and activator of transcription 3 (STAT3), a transcriptional activator implicated in fibrosis in endothelial cells, has been shown to induce EndMT via the TGF- β 1/ALK5/Smad4 signaling pathway [72]. In brain endothelial cells (BECs), activation of SNAIL and TAK1, which are regulators of EndMT, modulates the TGF- β 1 and interleukin 1- β (IL-1 β) signaling pathways, both of

which are involved in EndMT and BBB dysfunction [55]. FGF is crucial for endothelial cell functions, including proliferation, migration, maintenance of vascular integrity, and regulation of cell identity [73]. FGF and TGF- β exert opposing effects on endothelial cell function [74]. The FGF receptor 1 (FGFR1)/MAP4K4 pathway induces TGF- β /Smad signaling and inhibits EndMT in endothelial cells, playing an important role in mitigating fibrotic vascular diseases associated with EndMT [75]. FGF2 controls TGF β 1 signaling by regulating the expression of TGF β receptor complex (ALK5, TGFBR2, SARA) [76]. FGF2 modulates TGF- β 1 signaling by regulating the expression of the TGF- β receptor complex (ALK5, TGFBR2, SARA) [76]. VEGF, a key regulator of endothelial cell proliferation, differentiation, vascular permeability, and angiogenesis [77] may act as one of the most effective inhibitors of EndMT. VEGF exhibits an antifibrotic effect by inhibiting EndMT through suppression of TGF- β signaling and Smad2 activity [78].

EndMT in BECs is a process that contributes to atherosclerosis and BBB dysfunction by increasing BBB permeability, facilitating the filtration of immune substances, and promoting plaque formation [20]. Several studies have sought to elucidate the relationship between changes in junctional proteins in BECs and their impact on BBB permeability, atherosclerosis, and BBB breakdown. Claudin-5 (CLDN5), a critical tight junction protein, plays a vital role in BBB integrity, and its aberrant expression has been linked to neurodegenerative disorders [79]. One study demonstrated that CLDN5 deletion in the mouse brain resulted in increased intercellular leakage of molecules from the blood into the brain [80]. In adult mice, decreased CLDN5 expression selectively enhanced brain leakage depending on the size of the molecule and induced inflammatory responses in endothelial and glial cells lacking CLDN5 [80]. The regulation of BBB and neuronal function following ischemic stroke has been studied using occludin-deficient mice [62]. In a thrombosis mouse model, decreased expression of occludin was observed in BECs at ischemic lesions, leading to increased BBB permeability, accelerated brain tissue damage, and reduced angiogenesis following cerebral infarction, compared to control mice. Alongside the reduction in occludin expression after stroke, decreased levels of CLDN5 and ZO-1 were also noted. It remains unclear whether the absence of occludin directly reduced CLDN5 and ZO-1 expression or whether this effect was secondary to BBB disruption following stroke [62]. Nevertheless, occludin plays a

crucial role in maintaining BBB integrity in BECs and may present a potential therapeutic target for ischemic stroke.

Additionally, inflammatory responses not only affect the expression of junctional proteins but also alter the inflammatory phenotype of BECs. Inflammatory activation of BECs has been shown to increase the expression of adhesion molecules such as VCAM-1, ICAM-1, PECAM-1, and other adhesion molecules [64,81]. These changes play a significant role in the recruitment and migration of leukocytes to the brain, contributing to brain atherosclerosis [82,83]. Furthermore, BECs respond to inflammatory cytokines, including IL-1 β , interleukin-6 (IL-6), and TNF- α [84] as well as chemokines such as C-C motif chemokine ligand 2 (CCL2, MCP-1), CCL3, CCL5, C-X-C motif chemokine ligand 8 (CXCL8), CXCL10, and CXCL12, which recruit leukocytes into the brain [85,86].

THERAPEUTIC STRATEGIES RELATED TO ENDOTHELIAL REPROGRAMMING IN INTRACRANIAL ATHEROSCLEROSIS

Endothelial reprogramming encompasses various phenotypic transitions of endothelial cells, such as EndMT.

While current therapeutic approaches for intracranial atherosclerosis include lipid-lowering agents, antiplatelet medications, antihypertensive drugs, and anti-inflammatory compounds [87-89], these interventions have limited efficacy in disease reversal as they neither eliminate existing atherosclerotic lesions nor completely halt disease progression. EndMT has been implicated in the progression of atherosclerosis, and studies suggest that inhibiting EndMT can mitigate the development of the disease. Specifically, TGF- β , a key mediator of EndMT, plays a crucial role in atherosclerosis pathogenesis. Therefore, targeting TGF- β signaling may significantly reduce both EndMT and atherosclerosis. TGF- β modulates endothelial cell function through the regulation of two active signaling pathways mediated by its receptors: the ALK5-Smad2/3 pathway and the ALK1-Smad1/5 pathway. TGF- β receptor activation induces the ALK1-Smad1/5 pathway and concurrently inhibits the ALK5-Smad2/3 pathway, thereby suppressing EndMT [90]. FGF signaling is critical for maintaining vascular integrity and the expression of vascular endothelial growth factor receptor 2 (VEGFR2) [91]. Inhibition of FGF signaling promotes TGF- β signaling and accelerates EndMT [91]. Conversely, FGF-1 exerts its effects by reducing TGF- β signaling and inhibiting EndMT through the MAPK/ERK signaling pathway [92]. The transcription factor ETS1 is a key regulator of EndMT, which

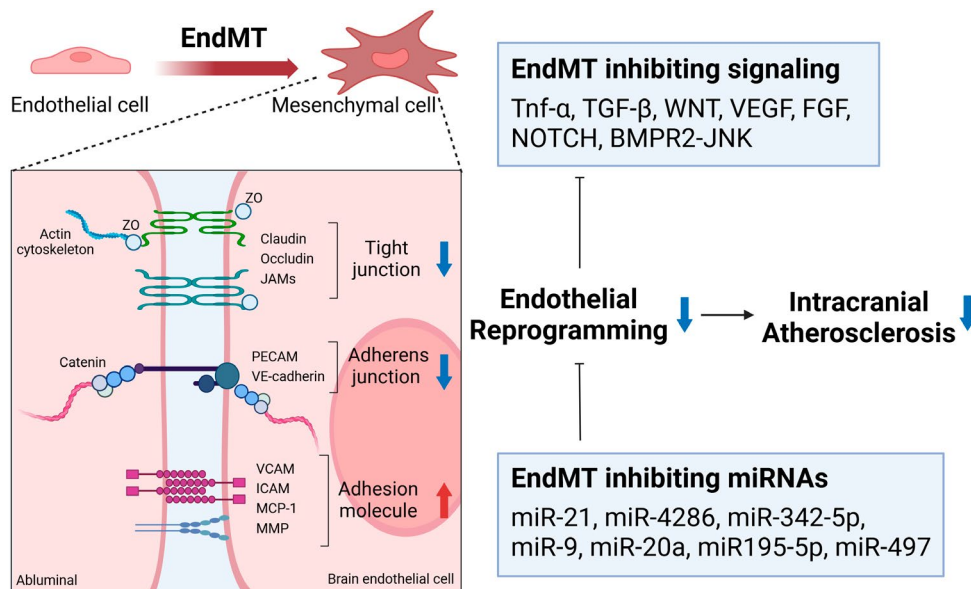


Fig. 2. Simplified scheme representing endothelial reprogramming in intracranial atherosclerosis. Schematic representation of tight junction (Claudin, occluding, JAMs), adherens junction (PECAM, VE-cadherin) and adhesion molecules (VCAM, ICAM, MCP-1, MMP) in the brain endothelial dysfunction. Inhibition of signaling and miRNAs that suppress endothelial reprogramming such as EndMT may reduce the progression of intracranial atherosclerosis.

is closely associated with BBB dysfunction. Under inflammatory conditions, inhibition of ETS1 exacerbates EndMT in central nervous system (CNS) endothelial cells [20]. Notably, upregulation of ETS1 in bEnd.3 cells treated with IL-1 β , an inflammatory cytokine, has been shown to inhibit EndMT and improve the function of BECs [93]. Furthermore, the expression of PIM1, a gene involved in atherosclerotic plaque formation [94], is upregulated in response to atherosclerosis. In human umbilical vein endothelial cells (HUVECs) subjected to EndMT, inhibition of PIM1 reduced mesenchymal marker expression and restored endothelial marker expression. Moreover, PIM1 knockdown in a mouse model of atherosclerotic plaques inhibited plaque progression by reducing the inflammatory response in endothelial cells and suppressing EndMT [94].

Numerous microRNAs (miRNAs) are known to regulate endothelial cell proliferation, migration, and the maintenance of vascular integrity by modulating transcription factors or participating in signaling pathways associated with EndMT [95]. The inhibition of EndMT-related signaling via miRNAs plays a pivotal role in preventing EndMT [96]. In endothelial cells, TGF- β enhances the expression of miR-21, which subsequently induces EndMT. Blocking miR-21 has been shown to attenuate TGF- β -induced EndMT through the PTEN/AKT signaling pathway [97]. In addition, miR-4286 expression is downregulated in endothelial cell dysfunction induced by (ox-LDL) [98]. Overexpression of miR-4286 diminishes TGF- β 1/Smad3 signaling, thereby alleviating endothelial cell damage [98]. The combination of FGF2 reduction and TGF- β 1 activation decreases miR-20a expression and activates Smad2/3, which increases EndMT [76]. Therefore, miR-20a expression can inhibit TGF- β 1 signaling by decreasing ALK5, TGFBR2 and SARA [76]. Inhibition of miR-9 increases MALAT1 expression and increases sensitivity to TGF- β and NF- κ B signaling, thereby activating inflammatory responses [99]. Inhibition of miR-9 can suppress glucose-induced EndMT by regulating TGFBR2, NF- κ B and MALAT1 signaling [99]. Inhibition of miR-195-5p can block EndMT by inhibiting TGF- β 1/Smad signaling [100]. Increased miR-497 targeting ROCK1 and ROCK2 reduced TGF- β 2-stimulated α -SMA and Snail levels [101]. Therefore, regulation of miR-497/ROCK signaling may attenuate EndMT [101] (Fig. 2). BBB dysfunction is a critical hallmark of intracranial atherosclerosis. Understanding the molecular mechanisms underlying BEC dysfunction during the pathological progression of intracra-

nial atherosclerosis could provide insights for developing potential therapeutic strategies aimed at restoring BBB function and mitigating the effects of intracranial atherosclerosis (Fig. 2).

CONCLUSION

In conclusion, this review examines the mechanistic relationship between endothelial reprogramming and intracranial atherosclerosis. Brain endothelial cells constitute a critical component of the BBB, regulating selective molecular transport between the circulation and central nervous system. EndMT promotes atherosclerosis progression through dual mechanisms: disruption of brain endothelial cell function and enhancement of atherosclerotic plaque formation. Thus, therapeutic strategies targeting EndMT may represent a promising approach for attenuating atherosclerosis development. Preclinical studies have demonstrated that modulation of EndMT-associated molecular pathways can mitigate various pathophysiological manifestations of atherosclerosis. However, comprehensive elucidation of the precise mechanisms governing endothelial cell reprogramming remains incomplete. Further investigations utilizing both in vivo models and cellular systems are essential to delineate these regulatory pathways and develop targeted therapeutic interventions.

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ABBREVIATIONS

Intracranial atherosclerosis (ICAS)
 blood-brain barrier (BBB)
 endothelial-to-mesenchymal transition (EndMT)
 brain endothelial cells (BECs)
 cell adhesion molecules (CAMs)
 reactive oxygen species (ROS)
 low-density lipoprotein (LDL)
 oxidized low-density lipoprotein (ox-LDL)
 nitric oxide (NO)
 nitric oxide synthase (NOS)

endothelial nitric oxide synthase (eNOS)
 neuronal nitric oxide synthase (nNOS)
 inducible nitric oxide synthase (iNOS)
 bilateral common carotid artery stenosis (BCAS)
 zonula occludens-1 (ZO-1)
 junctional adhesion molecules (JAM-A, JAM-B, JAM-C)
 platelet endothelial cell adhesion molecule-1 (PECAM-1)
 vascular cell adhesion molecule-1 (VCAM-1)
 matrix metalloproteinase-3 (MMP3)
 intracellular adhesion molecule-1 (ICAM-1)
 monocyte chemoattractant protein-1 (MCP-1)
 tumor necrosis factor-alpha (TNF- α)
 transforming growth factor-beta (TGF- β)
 vascular endothelial growth factor (VEGF)
 fibroblast growth factor (FGF)
 activin receptor-like kinases 1 and 5 (ALK1 and ALK5)
 signal transducer and activator of transcription 3 (STAT3)
 interleukin 1-beta (IL-1 β)
 FGF receptor 1 (FGFR1)
 Claudin-5 (CLDN5)
 interleukin-6 (IL-6)
 C-C motif chemokine ligand 2 (CCL2)
 C-X-C motif chemokine ligand 8 (CXCL8)
 vascular endothelial growth factor receptor 2 (VEGFR2)
 central nervous system (CNS)
 human umbilical vein endothelial cells (HUVECs)
 microRNAs (miRNAs)
 transforming growth factor beta receptor 2 (TGFBR2)
 Smad anchor for receptor activation (SARA)
 Metastasis associated lung adenocarcinoma transcript 1 (MALAT1)
 Rho-associated kinase 1 and 2 (ROCK1 and ROCK2)

REFERENCES

- Banerjee C, Chimowitz MI. Stroke caused by atherosclerosis of the major intracranial arteries. *Circ Res.* 2017;120:502-13.
- Chen LH, Spagnolo-Allende A, Yang D, Qiao Y, Gutierrez J. Epidemiology, pathophysiology, and imaging of atherosclerotic intracranial disease. *Stroke.* 2024;55:311-23.
- Ahmed R, Maqsood H, Bains RS, Gulraiz A, Kamal M. Intracranial atherosclerotic disease: current management strategies. *Ann Med Surg (Lond).* 2023;85:4903-8.
- Fujiyoshi A, Suri MFK, Alonso A, Selvin E, Chu H, Guallar E, et al. Hyperglycemia, duration of diabetes, and intracranial atherosclerotic stenosis by magnetic resonance angiography: the ARIC-NCS study. *J Diabetes Complications.* 2020;34:107605.
- Ma YH, Leng XY, Dong Y, Xu W, Cao XP, Ji X, et al. Risk factors for intracranial atherosclerosis: a systematic review and meta-analysis. *Atherosclerosis.* 2019;281:71-7.
- Liu R, Shao J. Research progress on risk factors related to intracranial artery, carotid artery, and coronary artery stenosis. *Front Cardiovasc Med.* 2022;9:970476.
- Ritz K, Denswil NP, Stam OC, van Lieshout JJ, Daemen MJ. Cause and mechanisms of intracranial atherosclerosis. *Circulation.* 2014;130:1407-14.
- Yang J, Zhang Y, Xue J, Guo Y, Liu S, Yao Y, et al. Hemodynamic effects of stenosis with varying severity in different segments of the carotid artery using computational fluid dynamics. *Sci Rep.* 2025;15:4896.
- Ajoolabady A, Pratico D, Lin L, Mantzoros CS, Bahijri S, Tuomilehto J, et al. Inflammation in atherosclerosis: pathophysiology and mechanisms. *Cell Death Dis.* 2024;15:817.
- Kim HJ, Lee EJ, Jung SH, Lee JW, Kim JS, Kim JB, et al. Cerebral atherosclerosis and early ischemic stroke after left-sided valve replacement surgery. *J Thorac Cardiovasc Surg.* 2022;163:967-76.e6.
- Trimm E, Red-Horse K. Vascular endothelial cell development and diversity. *Nat Rev Cardiol.* 2023;20:197-210.
- Berenji Ardestani S, Eftedal I, Pedersen M, Jeppesen PB, Norregaard R, Matchkov VV. Endothelial dysfunction in small arteries and early signs of atherosclerosis in ApoE knockout rats. *Sci Rep.* 2020;10:15296.
- Chen MB, Yang AC, Yousef H, Lee D, Chen W, Schaum N, et al. Brain endothelial cells are exquisite sensors of age-related circulatory cues. *Cell Rep.* 2020;30:4418-32.e4.
- de la Riva P, Marta-Enguita J, Rodriguez-Antiguedad J, Bergareche A, de Munain AL. Understanding endothelial dysfunction and its role in ischemic stroke after the outbreak of recanalization therapies. *Int J Mol Sci.* 2024;25.
- Nair AL, Groenendijk L, Overvest R, Fowke TM, Annida R, Mocellin O, et al. Human BBB-on-a-chip reveals barrier disruption, endothelial inflammation, and T cell migration under neuroinflammatory conditions. *Front Mol Neurosci.* 2023;16:1250123.
- Silva BR, Pernomian L, Bendhack LM. Contribution of oxidative stress to endothelial dysfunction in hypertension. *Front Physiol.* 2012;3:441.
- Cho JG, Lee A, Chang W, Lee MS, Kim J. Endothelial to mesenchymal transition represents a key link in the interaction between inflammation and endothelial dysfunction. *Front Immunol.* 2018;9:294.
- Singh A, Bhatt KS, Nguyen HC, Frisbee JC, Singh KK. Endothelial-to-mesenchymal transition in cardiovascular pathophysiology. *Int J Mol Sci.* 2024;25.

19. Zhang Z, Guo Q, Ma C, Zhao Z, Shi Q, Yu H, et al. USF1 transcriptionally activates USP14 to drive atherosclerosis by promoting EndMT through NLRC5/Smad2/3 axis. *Mol Med*. 2024;30:32.
20. Luo Y, Yang H, Wan Y, Yang S, Wu J, Chen S, et al. Endothelial ETS1 inhibition exacerbate blood-brain barrier dysfunction in multiple sclerosis through inducing endothelial-to-mesenchymal transition. *Cell Death Dis*. 2022;13:462.
21. Ohbuchi M, Shibuta M, Tetsuka K, Sasaki-Iwaoka H, Oishi M, Shimizu F, et al. Modeling of blood-brain barrier (BBB) dysfunction and immune cell migration using human BBB-on-a-chip for drug discovery research. *Int J Mol Sci*. 2024;25.
22. Sandoo A, van Zanten JJ, Metsios GS, Carroll D, Kitas GD. The endothelium and its role in regulating vascular tone. *Open Cardiovasc Med J*. 2010;4:302-12.
23. Brouns SLN, Provenzale I, van Geffen JP, van der Meijden PEJ, Heemskerk JWM. Localized endothelial-based control of platelet aggregation and coagulation under flow: a proof-of-principle vessel-on-a-chip study. *J Thromb Haemost*. 2020;18:931-41.
24. White TA, Johnson T, Zarzhevsky N, Tom C, Delacroix S, Holroyd EW, et al. Endothelial-derived tissue factor pathway inhibitor regulates arterial thrombosis but is not required for development or hemostasis. *Blood*. 2010;116:1787-94.
25. Cong X, Kong W. Endothelial tight junctions and their regulatory signaling pathways in vascular homeostasis and disease. *Cell Signal*. 2020;66:109485.
26. Mai J, Virtue A, Shen J, Wang H, Yang XF. An evolving new paradigm: endothelial cells--conditional innate immune cells. *J Hematol Oncol*. 2013;6:61.
27. Liao JK. Linking endothelial dysfunction with endothelial cell activation. *J Clin Invest*. 2013;123:540-1.
28. Janaszak-Jasiecka A, Ploska A, Wieronska JM, Dobrucki LW, Kalinowski L. Endothelial dysfunction due to eNOS uncoupling: molecular mechanisms as potential therapeutic targets. *Cell Mol Biol Lett*. 2023;28:21.
29. Sun L, Huang N, Yang C, Feng J, Chen H, Feng W, et al. Hirudin-based treatment of diabetes-induced erectile dysfunction through inhibition of the HIF-1 α to regulate RhoA/ROCK signaling pathway: an in vivo animal experiment. *Am J Mens Health*. 2025;19:15579883241310763.
30. Xia C, Hu J, Zhou K, Li Y, Yuan S, Li Q. Theoretical and experimental studies of the dynamic damage of endothelial cellular networks under ultrasound cavitation. *Cell Mol Bioeng*. 2025;18:15-28.
31. Benincasa G, Coscioni E, Napoli C. Cardiovascular risk factors and molecular routes underlying endothelial dysfunction: novel opportunities for primary prevention. *Biochem Pharmacol*. 2022;202:115108.
32. Pong S, Karmacharya R, Sofman M, Bishop JR, Lizano P. The role of brain microvascular endothelial cell and blood-brain barrier dysfunction in schizophrenia. *Complex Psychiatry*. 2020;6:30-46.
33. Tran KA, Zhang X, Predescu D, Huang X, Machado RF, Gothert JR, et al. Endothelial beta-catenin signaling is required for maintaining adult blood-brain barrier integrity and central nervous system homeostasis. *Circulation*. 2016;133:177-86.
34. Lippmann ES, Azarin SM, Kay JE, Nessler RA, Wilson HK, Al-Ahmad A, et al. Derivation of blood-brain barrier endothelial cells from human pluripotent stem cells. *Nat Biotechnol*. 2012;30:783-91.
35. Hamilos M, Petousis S, Parthenakis F. Interaction between platelets and endothelium: from pathophysiology to new therapeutic options. *Cardiovasc Diagn Ther*. 2018;8:568-80.
36. Pan X, Liu J, Zhong L, Zhang Y, Liu C, Gao J, et al. Identification of lipid metabolism-related biomarkers for diagnosis and molecular classification of atherosclerosis. *Lipids Health Dis*. 2023;22:96.
37. Duan H, Song P, Li R, Su H, He L. Attenuating lipid metabolism in atherosclerosis: the potential role of Anti-oxidative effects on low-density lipoprotein of herbal medicines. *Front Pharmacol*. 2023;14:1161657.
38. Ma Y, Huang Z, Zhou Z, He X, Wang Y, Meng C, et al. A novel antioxidant Mito-Tempol inhibits ox-LDL-induced foam cell formation through restoration of autophagy flux. *Free Radic Biol Med*. 2018;129:463-72.
39. Jiang H, Zhou Y, Nabavi SM, Sahebkar A, Little PJ, Xu S, et al. Mechanisms of oxidized LDL-mediated endothelial dysfunction and its consequences for the development of atherosclerosis. *Front Cardiovasc Med*. 2022;9:925923.
40. Wraith KS, Magwenzi S, Aburima A, Wen Y, Leake D, Naseem KM. Oxidized low-density lipoproteins induce rapid platelet activation and shape change through tyrosine kinase and Rho kinase-signaling pathways. *Blood*. 2013;122:580-9.
41. Liu J, Ren Y, Kang L, Zhang L. Oxidized low-density lipoprotein increases the proliferation and migration of human coronary artery smooth muscle cells through the upregulation of osteopontin. *Int J Mol Med*. 2014;33:1341-7.
42. Tousoulis D, Kampoli AM, Tentolouris C, Papageorgiou N, Stefanadis C. The role of nitric oxide on endothelial function. *Curr Vasc Pharmacol*. 2012;10:4-18.
43. Siragusa M, Thole J, Bibli SI, Luck B, Loot AE, de Silva K, et al. Nitric oxide maintains endothelial redox homeostasis through PKM2 inhibition. *EMBO J*. 2019;38:e100938.
44. Topf HG, Rauh M, Rascher W, Dotsch J, Klinge JM. Endothelial cells influence the sodium nitroprusside mediated inhibition of platelet aggregation by an as yet unknown pathway. *Thromb J*. 2012;10:6.
45. Sun HJ, Wu ZY, Nie XW, Bian JS. Role of endothelial dysfunction in cardiovascular diseases: the link between inflam-

- mation and hydrogen sulfide. *Front Pharmacol.* 2019;10:1568.
46. Arachchillage DJ, Mobayen G, Laffan M, Randi AM, Ahnstrom J, Pericleous C. A cell-based model to study mechanisms of endothelial-dependent thrombin generation in response to inflammation and its modulation by hydroxychloroquine. *Res Pract Thromb Haemost.* 2025;9:102665.
 47. Gao F, Lucke-Wold BP, Li X, Logsdon AF, Xu LC, Xu S, et al. Reduction of endothelial nitric oxide increases the adhesiveness of constitutive endothelial membrane ICAM-1 through Src-mediated phosphorylation. *Front Physiol.* 2017;8:1124.
 48. Zhou L, Zhu DY. Neuronal nitric oxide synthase: structure, subcellular localization, regulation, and clinical implications. *Nitric Oxide.* 2009;20:223-30.
 49. Tran N, Garcia T, Aniq M, Ali S, Ally A, Nauli SM. Endothelial nitric oxide synthase (eNOS) and the cardiovascular system: in physiology and in disease states. *Am J Biomed Sci Res.* 2022;15:153-77.
 50. Tan XL, Xue YQ, Ma T, Wang X, Li JJ, Lan L, et al. Partial eNOS deficiency causes spontaneous thrombotic cerebral infarction, amyloid angiopathy and cognitive impairment. *Mol Neurodegener.* 2015;10:24.
 51. Chen J, Zacharek A, Zhang C, Jiang H, Li Y, Roberts C, et al. Endothelial nitric oxide synthase regulates brain-derived neurotrophic factor expression and neurogenesis after stroke in mice. *J Neurosci.* 2005;25:2366-75.
 52. Asproni P, Cozzi A, Verin R, Lafont-Lecuelle C, Bienboire-Frosini C, Poli A, et al. Pathology and behaviour in feline medicine: investigating the link between vomeronasalitis and aggression. *J Feline Med Surg.* 2016;18:997-1002.
 53. An L, Shen Y, Chopp M, Zacharek A, Venkat P, Chen Z, et al. Deficiency of endothelial nitric oxide synthase (eNOS) exacerbates brain damage and cognitive deficit in a mouse model of vascular dementia. *Aging Dis.* 2021;12:732-46.
 54. Santhanam AV, d'Uscio LV, Smith LA, Katusic ZS. Uncoupling of eNOS causes superoxide anion production and impairs NO signaling in the cerebral microvessels of hph-1 mice. *J Neurochem.* 2012;122:1211-8.
 55. Derada Troletti C, Fontijn RD, Gowing E, Charabati M, van Het Hof B, Didouh I, et al. Inflammation-induced endothelial to mesenchymal transition promotes brain endothelial cell dysfunction and occurs during multiple sclerosis pathophysiology. *Cell Death Dis.* 2019;10:45.
 56. Yuan Y, Sun J, Dong Q, Cui M. Blood-brain barrier endothelial cells in neurodegenerative diseases: Signals from the "barrier". *Front Neurosci.* 2023;17:1047778.
 57. Schweiger B, Kievit FM. Glioblastoma induced blood-brain barrier dysfunction via a paracrine mechanism that increases claudin-1 expression. *Exp Brain Res.* 2025;243:70.
 58. Roher AE, Tyas SL, Maarouf CL, Dausgs ID, Kokjohn TA, Emmerling MR, et al. Intracranial atherosclerosis as a contributing factor to Alzheimer's disease dementia. *Alzheimers Dement.* 2011;7:436-44.
 59. Tayler H, Miners JS, Guzel O, MacLachlan R, Love S. Mediators of cerebral hypoperfusion and blood-brain barrier leakage in Alzheimer's disease, vascular dementia and mixed dementia. *Brain Pathol.* 2021;31:e12935.
 60. Li M, Li Y, Zuo L, Hu W, Jiang T. Increase of blood-brain barrier leakage is related to cognitive decline in vascular mild cognitive impairment. *BMC Neurol.* 2021;21:159.
 61. Derada Troletti C, de Goede P, Kamermans A, de Vries HE. Molecular alterations of the blood-brain barrier under inflammatory conditions: the role of endothelial to mesenchymal transition. *Biochim Biophys Acta.* 2016;1862:452-60.
 62. Sugiyama S, Sasaki T, Tanaka H, Yan H, Ikegami T, Kanki H, et al. The tight junction protein occludin modulates blood-brain barrier integrity and neurological function after ischemic stroke in mice. *Sci Rep.* 2023;13:2892.
 63. Olejarz W, Lacheta D, Kubiak-Tomaszewska G. Matrix metalloproteinases as biomarkers of atherosclerotic plaque instability. *Int J Mol Sci.* 2020;21.
 64. Zhang H, Shang J, Li W, Gao D, Zhang J. Increased expression of VCAM1 on brain endothelial cells drives blood-brain barrier impairment following chronic cerebral hypoperfusion. *ACS Chem Neurosci.* 2024;15:2028-41.
 65. Sanchez-Duffhues G, Garcia de Vinuesa A, van de Pol V, Geerts ME, de Vries MR, Janson SG, et al. Inflammation induces endothelial-to-mesenchymal transition and promotes vascular calcification through downregulation of BMPR2. *J Pathol.* 2019;247:333-46.
 66. Aisagbonhi O, Rai M, Ryzhov S, Atria N, Feoktistov I, Hatzopoulos AK. Experimental myocardial infarction triggers canonical Wnt signaling and endothelial-to-mesenchymal transition. *Dis Model Mech.* 2011;4:469-83.
 67. Guan G, Xie J, Dai Y, Han H. TFPI2 suppresses the interaction of TGF-beta2 pathway regulators to promote endothelial-mesenchymal transition in diabetic nephropathy. *J Biol Chem.* 2022;298:101725.
 68. Ma J, Sanchez-Duffhues G, Goumans MJ, Ten Dijke P. TGF-beta-induced endothelial to mesenchymal transition in disease and tissue engineering. *Front Cell Dev Biol.* 2020;8:260.
 69. Goumans MJ, Valdimarsdottir G, Itoh S, Lebrin F, Larsson J, Mummery C, et al. Activin receptor-like kinase (ALK)1 is an antagonistic mediator of lateral TGFbeta/ALK5 signaling. *Mol Cell.* 2003;12:817-28.
 70. van Meeteren LA, ten Dijke P. Regulation of endothelial cell plasticity by TGF-beta. *Cell Tissue Res.* 2012;347:177-86.
 71. Cooley BC, Nevado J, Mellad J, Yang D, St Hilaire C, Negro A, et al. TGF-beta signaling mediates endothelial-to-mesen-

- chymal transition (EndMT) during vein graft remodeling. *Sci Transl Med.* 2014;6:227ra34.
72. Becerra A, Rojas M, Vallejos A, Villegas V, Perez L, Cabello-Verrugio C, et al. Endothelial fibrosis induced by suppressed STAT3 expression mediated by signaling involving the TGF-beta1/ALK5/Smad pathway. *Lab Invest.* 2017;97:1033-46.
 73. Murakami M, Nguyen LT, Hatanaka K, Schachterle W, Chen PY, Zhuang ZW, et al. FGF-dependent regulation of VEGF receptor 2 expression in mice. *J Clin Invest.* 2011;121:2668-78.
 74. Chen PY, Simons M. Fibroblast growth factor-transforming growth factor beta dialogues, endothelial cell to mesenchymal transition, and atherosclerosis. *Curr Opin Lipidol.* 2018;29:397-403.
 75. Li J, Shi S, Srivastava SP, Kitada M, Nagai T, Nitta K, et al. FGFR1 is critical for the anti-endothelial mesenchymal transition effect of N-acetyl-seryl-aspartyl-lysyl-proline via induction of the MAP4K4 pathway. *Cell Death Dis.* 2017;8:e2965.
 76. Correia AC, Moonen JR, Brinker MG, Krenning G. FGF2 inhibits endothelial-mesenchymal transition through micro RNA-20a-mediated repression of canonical TGF-beta signaling. *J Cell Sci.* 2016;129:569-79.
 77. Bates DO. Vascular endothelial growth factors and vascular permeability. *Cardiovasc Res.* 2010;87:262-71.
 78. Illigens BM, Casar Berazaluce A, Poutias D, Gasser R, Del Nido PJ, Friehs I. Vascular endothelial growth factor prevents endothelial-to-mesenchymal transition in hypertrophy. *Ann Thorac Surg.* 2017;104:932-9.
 79. Greene C, Hanley N, Campbell M. Claudin-5: gatekeeper of neurological function. *Fluids Barriers CNS.* 2019;16:3.
 80. Vazquez-Liebanas E, Mocci G, Li W, Lavina B, Reddy A, O'Connor C, et al. Mosaic deletion of claudin-5 reveals rapid non-cell-autonomous consequences of blood-brain barrier leakage. *Cell Rep.* 2024;43:113911.
 81. Kalinowska A, Losy J. PECAM-1, a key player in neuroinflammation. *Eur J Neurol.* 2006;13:1284-90.
 82. Singh V, Kaur R, Kumari P, Pasricha C, Singh R. ICAM-1 and VCAM-1: gatekeepers in various inflammatory and cardiovascular disorders. *Clin Chim Acta.* 2023;548:117487.
 83. Winneberger J, Schols S, Lessmann K, Randez-Garbayo J, Bauer AT, Mohamud Yusuf A, et al. Platelet endothelial cell adhesion molecule-1 is a gatekeeper of neutrophil transendothelial migration in ischemic stroke. *Brain Behav Immun.* 2021;93:277-87.
 84. O'Carroll SJ, Kho DT, Wiltshire R, Nelson V, Rotimi O, Johnson R, et al. Pro-inflammatory TNFalpha and IL-1beta differentially regulate the inflammatory phenotype of brain microvascular endothelial cells. *J Neuroinflammation.* 2015;12:131.
 85. Chui R, Dorovini-Zis K. Regulation of CCL2 and CCL3 expression in human brain endothelial cells by cytokines and lipopolysaccharide. *J Neuroinflammation.* 2010;7:1.
 86. Subileau EA, Rezaie P, Davies HA, Colyer FM, Greenwood J, Male DK, et al. Expression of chemokines and their receptors by human brain endothelium: implications for multiple sclerosis. *J Neuropathol Exp Neurol.* 2009;68:227-40.
 87. Wabnitz AM, Turan TN. Optimal medical management of atherosclerotic intracranial stenosis. *Stroke.* 2024;55:335-43.
 88. Kim JS. Role of blood lipid levels and lipid-lowering therapy in stroke patients with different levels of cerebral artery diseases: reconsidering recent stroke guidelines. *J Stroke.* 2021;23:149-61.
 89. Kim JS, Bang OY. Medical treatment of intracranial atherosclerosis: an update. *J Stroke.* 2017;19:261-70.
 90. Lebrin F, Goumans MJ, Jonker L, Carvalho RL, Valdimarsdottir G, Thorikay M, et al. Endoglin promotes endothelial cell proliferation and TGF-beta/ALK1 signal transduction. *EMBO J.* 2004;23:4018-28.
 91. Chen PY, Qin L, Barnes C, Charisse K, Yi T, Zhang X, et al. FGF regulates TGF-beta signaling and endothelial-to-mesenchymal transition via control of let-7 miRNA expression. *Cell Rep.* 2012;2:1684-96.
 92. Ramos C, Becerril C, Montano M, Garcia-De-Alba C, Ramirez R, Checa M, et al. FGF-1 reverts epithelial-mesenchymal transition induced by TGF-beta1 through MAPK/ERK kinase pathway. *Am J Physiol Lung Cell Mol Physiol.* 2010;299:L222-31.
 93. Lu Q, Ma J, Zhao Y, Ding G, Wang Y, Qiao X, et al. Disruption of blood-brain barrier and endothelial-to-mesenchymal transition are attenuated by Astragalus polysaccharides mediated through upregulation of ETS1 expression in experimental autoimmune encephalomyelitis. *Biomed Pharmacother.* 2024;180:117521.
 94. Xue Z, Han M, Sun T, Wu Y, Xing W, Mu F, et al. PIM1 instigates endothelial-to-mesenchymal transition to aggravate atherosclerosis. *Theranostics.* 2025;15:745-65.
 95. Chamorro-Jorganes A, Araldi E, Suarez Y. MicroRNAs as pharmacological targets in endothelial cell function and dysfunction. *Pharmacol Res.* 2013;75:15-27.
 96. Kim J. MicroRNAs as critical regulators of the endothelial to mesenchymal transition in vascular biology. *BMB Rep.* 2018;51:65-72.
 97. Kumarswamy R, Volkmann I, Jazbutyte V, Dangwal S, Park DH, Thum T. Transforming growth factor-beta-induced endothelial-to-mesenchymal transition is partly mediated by micro RNA-21. *Arterioscler Thromb Vasc Biol.* 2012;32:361-9.
 98. He Z, Xue H, Liu P, Han D, Xu L, Zeng X, et al. miR-4286/TGF-beta1/Smad3-negative feedback loop ameliorated vascular endothelial cell damage by attenuating apoptosis and inflammatory response. *J Cardiovasc Pharmacol.* 2020;75:

- 446-54.
99. Wang E, Feng B, Chakrabarti S. MicroRNA 9 is a regulator of endothelial to mesenchymal transition in diabetic retinopathy. *Invest Ophthalmol Vis Sci.* 2023;64:13.
100. Ding H, Yao J, Xie H, Wang C, Chen J, Wei K, et al. Micro RNA-195-5p downregulation inhibits endothelial mesenchymal transition and myocardial fibrosis in diabetic cardio-myopathy by targeting Smad7 and inhibiting transforming growth factor beta 1-Smads-snail pathway. *Front Physiol.* 2021;12:709123.
101. Liu F, Zhang S, Xu R, Gao S, Yin J. Melatonin attenuates endothelial-to-mesenchymal transition of glomerular endothelial cells via regulating miR-497/ROCK in diabetic nephropathy. *Kidney Blood Press Res.* 2018;43:1425-36.