

Effects of Mineral Trioxide Aggregate on the Proliferation and Differentiation of Human Dental Pulp Stromal Cells from Permanent and Deciduous Teeth

Seunghye Kim¹, Mijeong Jeon¹, Dong Min Shin², Jae Ho Lee¹, Je Seon Song¹

¹Department of Pediatric Dentistry, College of Dentistry and Oral Science Research Center, Yonsei University, Seoul, Korea

²Department of Oral Science, College of Dentistry, Yonsei University, Seoul, Korea

Abstract

Mineral trioxide aggregate (MTA) has recently been used as a pulpotomy medicament for primary molars. The aim of this study was to evaluate and compare the proliferation and differentiation potential of dental pulp stromal cells of permanent teeth and deciduous teeth cultured on MTA-coated surface. Human dental pulp stromal cells were obtained from human permanent premolars and deciduous teeth and cultured on MTA-coated culture plates. The cells were subjected to proliferation assay and cell cycle analysis. Their differentiation potential was evaluated by analysing changes in the mRNA expressions of runt-related transcriptional factor 2 (*Runx2*) and alkaline phosphatase (*ALP*). Morphological changes of cells in direct contact with MTA were observed using scanning electron microscopy (SEM). The proliferation rates, distribution of cell cycles and mRNA expression patterns of *Runx2* and *ALP* were similar in both types of pulpal cells. SEM observations revealed that both types changed into more dendrite-like cells. On the surface of MTA, human dental pulp stromal cells from deciduous and permanent teeth were able to both proliferate and differentiate into cells that induce mineralization. MTA is suitable as a biocompatible pulpotomy medicament for primary teeth.

Key words : Mineral Trioxide Aggregate (MTA), Human dental pulp stromal cells, Deciduous teeth, Permanent teeth, Proliferation, Differentiation

I. Introduction

Several pulp medicaments are commonly used for pulpotomy in primary molars, including formocresol and ferric sulphate. However, the use of formocresol is now decreasing in many countries as a result of its carcinogenic properties in human^{1,2)} and ferric sulphate is known to irritate the pulp tissue and provoke an inflammatory reaction in some cases³⁻⁶⁾.

Mineral trioxide aggregate (MTA) has recently been applied clinically with successful outcomes as an alterna-

tive pulp medicament for pulpotomy in primary molars^{7,8)}. Since it was introduced by Torabinejad *et al.* in 1993⁹⁾, MTA has been widely applied in various endodontic procedures, such as pulp capping, pulpotomy, apical barrier formation in immature permanent teeth with necrotic pulp, repair of root perforation, root-end filling and root canal filling. MTA is a biocompatible material, non-toxic to the host, and has been reported to induce dentin bridge formation and hard-tissue formation on exposed pulp tissue without pulpal necrosis in animal studies^{6,10)}.

Corresponding author : Je Seon Song

Department of Pediatric Dentistry, College of Dentistry, Yonsei University, 50 Yonsei-ro, Seodaemun-gu, Seoul, 120-752, Korea

Tel : +82-2-2228-3176 / Fax : +82-2-392-7420 / E-mail: SONGJS@yuhs.ac

Received November 30, 2012 / Revised June 4, 2013 / Accepted June 11, 2013

※ This study was supported by a grant of the Korea Healthcare Technology R&D Project, Ministry for Health, Welfare and Family Affairs, Republic of Korea (no. A100517) and Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (2011-0022160).

Previous *in vitro* research has demonstrated the biocompatibility of MTA using human dental pulp stromal cells from permanent teeth¹¹⁻¹³. However, few *in vitro* studies have investigated the pulpal reaction to MTA in deciduous teeth¹⁴. The aim of this study was to compare the viability and differentiation potential of pulpal cells from permanent and deciduous teeth in direct contact with MTA.

II. Materials and Methods

1. Preparation of MTA

White MTA (ProRoot MTA, Dentsply Tulsa Dental, Tulsa, OK, USA) was used in all experiments. The material was mixed according to the manufacturer's instructions and then plated in culture plates (BD Falcon, Franklin Lakes, NJ, USA) or on round glass slides (8 mm in diameter), as shown in Fig. 1. The MTA was left to set completely for 3~4 days at 37°C in a humidified 5%-CO₂ incubator. After complete setting, the MTA-coated plates and glass slides were sterilized under ethylene oxide gas.

2. Cell cultures

The experimental protocol was approved by the Institutional Review Board of the Dental Hospital, Yonsei University, and informed consent to participate was obtained from all of the subjects and their parents (#2-2010-0012). Human dental pulp tissue was obtained from healthy permanent premolars (*n* = 3; pa-



Fig. 1. White MTA coated onto 24-well culture plates. The first and third rows are coated with white MTA while the second and fourth rows are left uncoated for the control.

tients aged 13~18 years) that had been extracted for orthodontic purposes, and from deciduous teeth (*n* = 4; patients aged 2~7 years) from which mechanically exposed pulp tissue was extirpated during routine treatment for caries (Table 1). The extracted teeth and extirpated pulp tissue were stored in Dulbecco's Modified Eagle's Medium (Invitrogen, Carlsbad, CA, USA). The pulp tissue, corresponding to the middle one-third of the root, was separated and subjected to primary culture by outgrowth method. Briefly, the separated pulp tissue was minced into about 1.0 mm³ size and pressed gently unto 60 mm culture dish (BD Falcon, Franklin Lakes, NJ, USA) using cover glass (Superior, Lauda-Königshofen, Germany). The explants were cultured in alpha minimum essential medium (α -MEM; Invitrogen) supplemented with 10% fetal bovine serum (FBS; Invitrogen), 100 U/ml penicillin and 100 ug/ml streptomycin (Invitrogen) at 37°C in 5% CO₂. The isolated cells from 3 samples of premolars were blended at the first passage, and the third to fifth passages were used in the subsequent experiments. The same procedure was performed for the pulp stromal cells obtained from four samples of deciduous teeth.

3. Proliferation assay

The proliferation rate of human dental pulp cells from permanent teeth and deciduous teeth in direct contact with MTA was measured using Cell Counting Kit-8 (Dojindo, Kumamoto, Japan), according to the manufacturer's instructions. Human dental pulp cells were seeded at 5×10^3 cells/well in 24-well culture plates that were either uncoated or coated with MTA. After 1, 3, 5 and 7 days of culture period, the optical density of the yellow-coloured formazan formed by vital cells was measured at 450 nm using a Benchmark Plus Microplate spectrophotometer (Bio-Rad Laboratories, Hercules, CA, USA).

Table 1. Information about the source of extirpated pulp tissue regarding the tooth number, age, and gender of the subjects

Source	Tooth Number	Age	Gender
Permanent Teeth	#14	18Y	F
	#14	15Y	M
	#24	13Y 7M	M
Primary Teeth	#54	7Y 4M	M
	#53	5Y 3M	F
	#64	4Y 5M	F
	#74	2Y 2M	M

4. Cell cycle analysis

Cells were seeded at 5×10^5 cells/well onto MTA-coated or uncoated 60-mm culture plates, cultured for 3 days, and then harvested by trypsinization and fixed in cold 70% ethanol for 1 hour at 4°C. After washing twice with phosphate-buffered saline (Invitrogen), samples were incubated in 0.2 mg/ml RNase A (LaboPass, Sapporo, Japan) for 1 hour at 37°C and then stained with propidium iodide (40 µg/ml; #P-4170, Sigma) at 4°C for 30 min. Flow cytometric analysis was performed using a FACS Calibur flow cytometer (BD Biosciences, San Jose, CA, USA), and cell cycle analysis was conducted using FCS Express V3 software (De Novo Software, Los Angeles, CA, USA).

5. Gene expression analysis using the quantitative reverse-transcription polymerase chain reaction (RT-PCR)

Human dental pulp cells from permanent and deciduous teeth were seeded at a concentration of 1×10^5 cells/well in either MTA-coated or uncoated 12-well culture plates. They were harvested from the culture plates at 1, 2 and 4 days after seeding by trypsinization, and total cell RNA was extracted with the RNeasy Micro Kit (Qiagen, Valencia, CA, USA), according to the manufacturer's instructions. The concentration of RNA was calculated using a spectrophotometer (Nanodrop ND-1000, Thermo Scientific, Waltham, MA, USA). A 1-µg aliquot of each RNA sample was reverse transcribed into cDNA using an Maxime reverse transcription premix kit (Intron Biotechnology, Seoul, Korea). According to the manufacturer's instructions, the total RNA was reverse transcribed using an oligo d(T)15 primer at 45°C for 1 hour, and the reaction was stopped by incubation at 95°C for 5 min.

The complemented DNA was subjected to quantitative RT-PCR analysis using SYBR premix Ex Taq (Takara Bio, Otsu, Japan) and the Thermal Cycler Dice real-time system (Takara Bio), according to the manufacturer's instructions. Each PCR assay was performed with. The program was set at 95°C for 10 s for the initial denaturing step, followed by 45 cycles for denaturation at 95°C for 5 s, annealing at 60°C for 15 s and amplification at 72°C for 10 s. Amplification specificity was confirmed by visualizing the PCR products on 1.5% agarose gels and by melting-curve analysis (from 60°C to 95°C) after

the completion of 45 cycles. The values for each gene were normalized to the expression levels of glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*), and the relative quantities of the studied genes were calculated. The specific primers used in this study included *Runx2* (NM_004348.3: forward primer, 5'-CACTGGCGCTG-CAACAAGA-3'; reverse primer, 5'-CATTCCGGAGC-TCAGCAGAATAA-3'), alkaline phosphatase (ALP; NM_000478.4: forward primer, 5'-GGACCATTCC-CACGTCTTCAC-3'; reverse primer, 5'-CCTTGTAGCC-AGGCCCATTTG-3') and *GAPDH* (NM_002046.3: forward primer, 5'-TCCTGCACCACCAACTGCTT-3'; reverse primer, 5'-TGGCAGTGATGGCATGGAC-3').

6. Scanning electron microscopy (SEM) analysis

Human dental pulp cells from permanent and deciduous teeth were seeded at a concentration of 1×10^5 cells/well in four-well plates, whereby MTA-coated round glass slides were placed with the MTA facing upward. After 5 days, the pulp cells on the MTA-coated slides were incubated in fixative solution containing 2% paraformaldehyde, 2% glutaraldehyde and 0.5% CaCl₂, followed by post-fixation using 1% OsO₄ (all these chemicals were purchased from Sigma). After dehydration and drying, the samples were coated with a 100-nm layer of platinum and then visualized using a Hitachi S-3000N scanning electron microscope (Hitachi, Tokyo, Japan) at an accelerating voltage of 20.0 kV and a magnification of $\times 1500$.

7. Statistical analysis

All of the experiments were repeated under at least three independent conditions. All data are presented as mean and standard deviation values. Any significant differences between human dental pulp stromal cells from permanent teeth and deciduous teeth were determined using Student's *t*-test with SPSS software (version 17.0, SPSS, Chicago, IL, USA). The level of statistical significance was set at $p < 0.05$.

III. Results

1. Proliferation assay and cell cycle analysis

In the proliferation assay, there was no significant difference between pulpal cells from permanent teeth and

deciduous teeth when cultured on either an uncoated culture plate or on the MTA surface. However, the proliferation rate was significantly higher on the culture plate compared to the MTA-coated surface for both cell types (Fig. 2). The cell cycle analysis revealed similar percentages of cells in the G1 phase in all samples (63~64%; Fig. 3).

2. Quantitative RT-PCR

The expression of *Runx2* increased at day 2, and then gradually decreased thereafter, while that of *ALP* continued to gradually increase over the 7-day experimental period. However, these expression levels did not show significant difference between pulpal cells from permanent teeth and deciduous teeth cultured on an MTA-coated surface (Fig. 4).

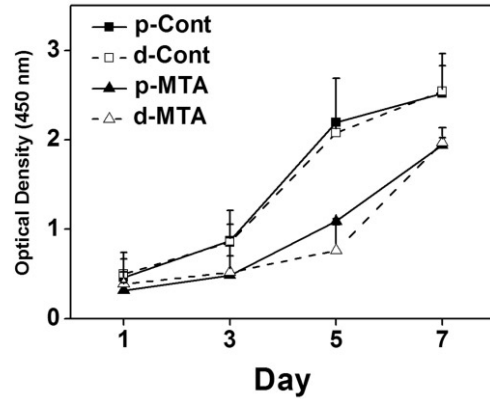


Fig. 2. Proliferation of pulpal cells from permanent and deciduous teeth in direct contact with MTA. The Y-axis indicates the optical density of yellow-coloured formazan converted by the cells. Abbreviations: p-Cont, pulpal cells from permanent teeth on a plate not coated with white MTA; d-Cont, pulpal cells from deciduous teeth on a plate not coated with white MTA; p-MTA, pulpal cells from permanent teeth on a plate coated with white MTA; d-MTA, pulpal cells from deciduous teeth on a plate not coated with white MTA.

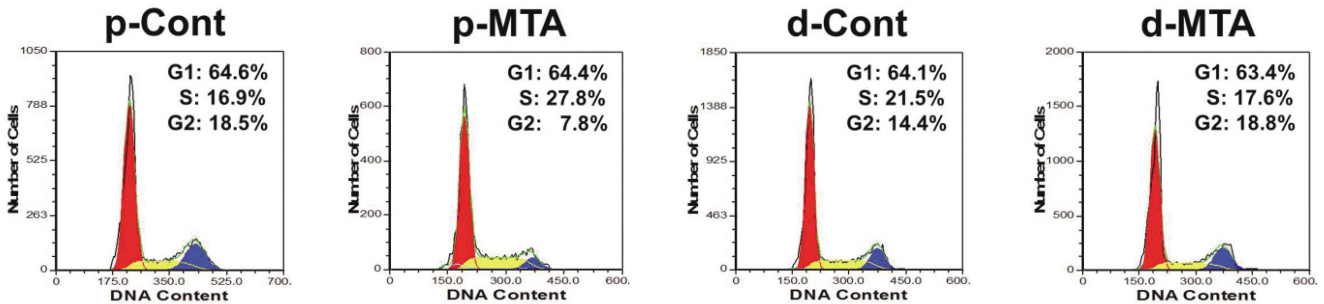


Fig. 3. Cell cycle distributions of pulpal cells on direct contact to MTA.

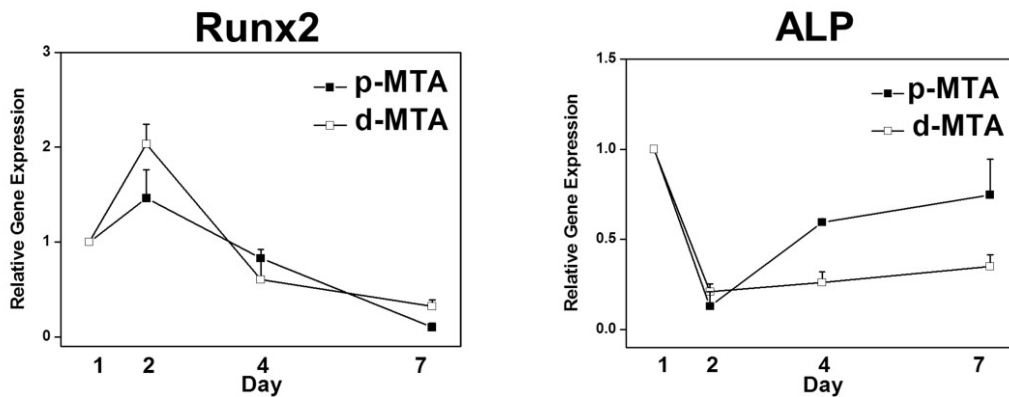


Fig. 4. Chronological changes in the gene expressions of *Runx2* and *ALP* of pulpal cells in direct contact with white MTA. Gene expression levels at 1 day after seeding were normalized to 1.

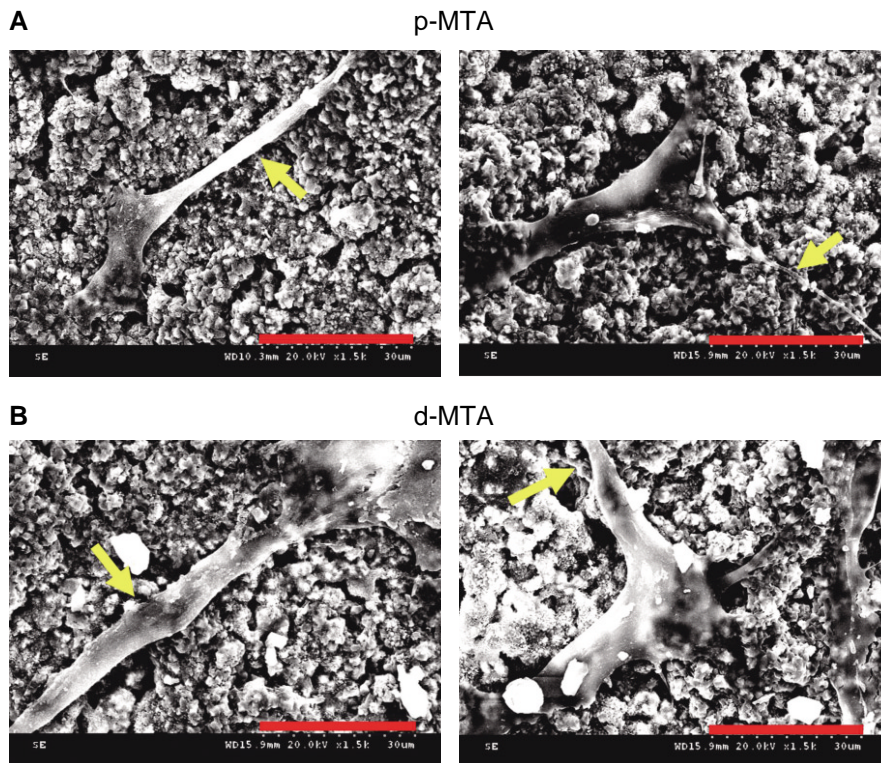


Fig. 5. SEM images of pulpal cells cultured on an MTA-coated surface from permanent (A) and deciduous (B) teeth. Arrows indicates dendritic process resembling odontoblast process. Scale bars: 30 μ m.

3. SEM analysis

Human pulpal cells from permanent teeth and deciduous teeth were able to attach to the MTA coating, presenting a more dendrite-like morphology with several branches, rather than a spindle shape (Fig. 5).

IV. Discussion

In previous studies, investigators have reported that dental pulp cells obtained through primary culture of dental-pulp tissue from permanent and shedding deciduous teeth present different features^{15,16}. The pulp stromal cells from permanent teeth are known to be capable of differentiating into odontoblasts and form a dentin-pulp-like complex following *in vivo* transplantation¹⁷. In contrast, those from exfoliated deciduous teeth fail to reconstitute a dentin-pulp-like complex *in vivo*, but do induce recipient cell-mediated bone formation *in vivo*¹⁵. In addition, the pulp cells from deciduous teeth exhibit a higher proliferation rate and increased cell-population doubling^{15,18}. Such differences found in previous studies sug-

gest that the pulpal reactions to MTA may differ between permanent and deciduous teeth.

In this study, unlike previous studies, the proliferation rates measured were similar in pulpal cells from permanent teeth and deciduous teeth, regardless of the presence of an MTA surface coating. While previous studies used dental pulp from exfoliated deciduous teeth, we used dental pulp extirpated from deciduous teeth with a sound root. Given that deciduous teeth go through various developmental stages from eruption through to shedding, the stage at which pulp tissue is obtained may be an important determinant of the pulpal characteristics, because pulp components and gene expression patterns may differ according to the stage.

The biocompatibility of MTA has been confirmed through many cell-culture studies, and it is known to be one of the least cytotoxic dental materials^{19,20}. In the present study, the proliferation rate of both cell types was reduced on MTA-coated culture plate, but the proportions of cells in the G1 phase were similar, regardless of the presence of the MTA coating. This indicates that the pulpal cells from permanent and deciduous teeth ex-

hibit similar viabilities on MTA, and confirms that MTA is a biocompatible material. The reduced proliferation rate on the MTA-coated surface can be explained by the surface roughness of set MTA. Previous studies that found an increased proliferation of pulp cells on MTA blocks used an insert-well system to prevent direct contact between the cells and the tested material^{11,21,22}. Direct contact between the test materials and the cells may introduce unfavourable factors that could obscure or inhibit the actual effect of the materials on cell survival and growth. In the present study, the MTA coating on the plate increased the roughness and irregularity of its surface, which may have inhibited the initial attachment of the cells, consequently leading to a reduction in actual numbers of seeded cells. However, the use of the insert-well system is limited in that it can only be used to measure the effects of the diffusible components of the material, while our system allowed the full effect of MTA in direct contact with the cells to be investigated.

It has been reported that MTA induces mineralization under both *in vitro* and *in vivo* conditions²³⁻²⁷. We have shown herein that the expression levels of *Runx2* and *ALP* of pulpal cells were up-regulated when they were cultured on an MTA-coated surface. *Runx2* is a transcriptional activator of osteogenesis-associated genes and is essential for osteoblast differentiation and bone formation²⁸. *Runx2* also regulates the proliferation and differentiation of odontoblasts and increases the expression of dentin sialophosphoprotein (*DSPP*) which is the main non-collagenous component of the odontoblastic extracellular matrix in immature odontoblasts^{29,30}. *ALP* is another important regulator during the mineralization of bone and reparative dentin³¹. *ALP* expression gradually increases during development of collagen matrix, which precedes calcium accumulation³². In this study, short expression of *Runx2* with its peak at Day 4 coincides with its character as the early marker initiating osteogenic or odontogenic differentiation. Gradual increase of *ALP* expression through 8-day period indicates development of collagen matrix. The SEM images showed that the deciduous and permanent pulp cells grown on an MTA block had dendritic process that resembled mature odontoblastic process. These findings indicate that human pulpal stromal cells from permanent and deciduous teeth may differentiate to induce hard-tissue formation when in direct contact with MTA. However, odontogenic differentiation was not confirmed in this study because we did not investigate either extracellular matrix

mineralization or the expression patterns of other mineralization-related genes, such as *DSPP*, osteocalcin, and osteopontin, which are expressed later in the mineralization process.

In conclusion, the results of the described experiments indicate that MTA is a biocompatible material for human pulpal stromal cells in both permanent and deciduous teeth. However, neither the differentiation pattern nor the mechanism of hard-tissue formation was clarified. Further studies are required to distinguish any difference in the mechanism of hard-tissue formation and composition of the final mineral product between permanent and deciduous teeth.

V. Conclusions

The present study investigated and compared the pulpal reactions of permanent and deciduous teeth when the pulp is in direct contact with set MTA *in vitro*. The proliferation of these cells was evaluated by (1) determining the relative numbers of vital cells and (2) cell cycle analysis. The differentiation of pulpal cells on MTA was evaluated using quantitative RT-PCR and by observation under SEM.

MTA did not interfere with the proliferation of human pulpal stromal cells from deciduous teeth as in permanent teeth. MTA induced the differentiation of human pulpal stromal cells from deciduous teeth into cells that form hard tissue, as it did in permanent teeth. This *in vitro* study confirmed MTA as a biocompatible pulp medicament for deciduous teeth, thereby providing biological support for its clinical use in deciduous teeth.

References

1. Judd PL, Kenny DJ: Formocresol concerns. A review. *J Can Dent Assoc*, 53:401-404, 1987.
2. International Agency for Research on Cancer: IARC classifies formaldehyde as carcinogenic to humans. Press release no. 153: Lyon: World Health Organization International Agency for research on Cancer, 2004.
3. Deery C: Formocresol and ferric sulfate have similar success rates in primary molar pulpotomy. In carious primary molars does a pulpotomy performed with ferric sulphate, compared with formocresol, result in greater clinical/radiographic success? *Evid Based Dent*, 6:70, 2005.
4. Fei AL, Udin RD, Johnson R: A clinical study of fer-

- ric sulfate as a pulpotomy agent in primary teeth. *Pediatr Dent*, 13:327-332, 1991.
5. Loh A, O'Hoy P, Tran X, *et al.*: Evidence-based assessment: evaluation of the formocresol versus ferric sulfate primary molar pulpotomy. *Pediatr Dent*, 26:401-409, 2004.
 6. Shayegan A, Petein M, Abbeele AV: Beta-tricalcium phosphate, white mineral trioxide aggregate, white Portland cement, ferric sulfate, and formocresol used as pulpotomy agents in primary pig teeth. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*, 105:536-542, 2008.
 7. Peng L, Ye L, Tan H, Zhou X: Evaluation of the formocresol versus mineral trioxide aggregate primary molar pulpotomy: a meta-analysis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*, 102:e40-44, 2006.
 8. Noorollahian H: Comparison of mineral trioxide aggregate and formocresol as pulp medicaments for pulpotomies in primary molars. *Br Dent J*, 204:E20, 2008.
 9. Torabinejad M, Watson TF, Pitt Ford TR: Sealing ability of a mineral trioxide aggregate when used as a root end filling material. *J Endod*, 19:591-595, 1993.
 10. Salako N, Joseph B, Ritwik P, *et al.*: Comparison of bioactive glass, mineral trioxide aggregate, ferric sulfate, and formocresol as pulpotomy agents in rat molar. *Dent Traumatol*, 19:314-320, 2003.
 11. Min KS, Yang SH, Kim EC: The combined effect of mineral trioxide aggregate and enamel matrix derivative on odontoblastic differentiation in human dental pulp cells. *J Endod*, 35:847-851, 2009.
 12. Paranjpe A, Cacalano NA, Hume WR, Jewett A: N-acetyl cysteine mediates protection from 2-hydroxyethyl methacrylate induced apoptosis via nuclear factor kappa B-dependent and independent pathways: potential involvement of JNK. *Toxicol Sci*, 108:356-366, 2009.
 13. Takita T, Hayashi M, Takeichi O, *et al.*: Effect of mineral trioxide aggregate on proliferation of cultured human dental pulp cells. *Int Endod J*, 39:415-422, 2006.
 14. Wang MY, Liu H, Li SL, Qin M: [Effects of mineral trioxide aggregate and calcium hydroxide on the proliferation and differentiation capacity of pulp cells of primary teeth]. *Zhonghua Kou Qiang Yi Xue Za Zhi*, 43:524-527, 2008.
 15. Miura M, Gronthos S, Zhao M, *et al.*: SHED: stem cells from human exfoliated deciduous teeth. *Proc Natl Acad Sci U S A*, 100:5807-5812, 2003.
 16. Gronthos S, Brahimi J, Li W, *et al.*: Stem cell properties of human dental pulp stem cells. *J Dent Res*, 81:531-535, 2002.
 17. Gronthos S, Mankani M, Brahimi J, *et al.*: Postnatal human dental pulp stem cells (DPSCs) in vitro and in vivo. *Proc Natl Acad Sci U S A*, 97:13625-13630, 2000.
 18. Nakamura S, Yamada Y, Katagiri W, *et al.*: Stem cell proliferation pathways comparison between human exfoliated deciduous teeth and dental pulp stem cells by gene expression profile from promising dental pulp. *J Endod*, 35:1536-1542, 2009.
 19. Torabinejad M, Parirokh M: Mineral trioxide aggregate: a comprehensive literature review--part II: leakage and biocompatibility investigations. *J Endod*, 36:190-202, 2010.
 20. Camilleri J, Pitt Ford TR: Mineral trioxide aggregate: a review of the constituents and biological properties of the material. *Int Endod J*, 39:747-754, 2006.
 21. Choi Y-S, Lee N-Y, Lee S-H: Gene expression of exposure to mineral trioxide aggregate (MTA) on dental pulp cells. *J Korean Acad Pediatr Dent*, 35:30-37, 2008.
 22. Moghaddame-Jafari S, Mantellini MG, Botero TM, *et al.*: Effect of ProRoot MTA on pulp cell apoptosis and proliferation in vitro. *J Endod*, 31:387-391, 2005.
 23. Ford TR, Torabinejad M, Abedi HR, *et al.*: Using mineral trioxide aggregate as a pulp-capping material. *J Am Dent Assoc*, 127:1491-1494, 1996.
 24. Faraco IM, Jr., Holland R: Response of the pulp of dogs to capping with mineral trioxide aggregate or a calcium hydroxide cement. *Dent Traumatol*, 17:163-166, 2001.
 25. Accorinte ML, Loguercio AD, Reis A, *et al.*: Response of human dental pulp capped with MTA and calcium hydroxide powder. *Oper Dent*, 33:488-495, 2008.
 26. Moretti AB, Sakai VT, Oliveira TM, *et al.*: The effectiveness of mineral trioxide aggregate, calcium hydroxide and formocresol for pulpotomies in primary teeth. *Int Endod J*, 41:547-555, 2008.
 27. Yasuda Y, Ogawa M, Arakawa T, *et al.*: The effect of mineral trioxide aggregate on the mineralization

- ability of rat dental pulp cells: an in vitro study. *J Endod*, 34:1057-1060, 2008.
28. Ducy P, Starbuck M, Priemel M, *et al.*: A Cbfa1-dependent genetic pathway controls bone formation beyond embryonic development. *Genes Dev*, 13:1025-1036, 1999.
29. Chen S, Rani S, Wu Y, *et al.*: Differential regulation of dentin sialophosphoprotein expression by Runx2 during odontoblast cytodifferentiation. *J Biol Chem*, 280:29717-29727, 2005.
30. Mizuno M, Imai T, Fujisawa R, *et al.*: Bone sialoprotein (BSP) is a crucial factor for the expression of osteoblastic phenotypes of bone marrow cells cultured on type I collagen matrix. *Calcif Tissue Int*, 66:388-396, 2000.
31. Butler WT: Dentin matrix proteins and dentinogenesis. *Connect Tissue Res*, 33:59-65, 1995.
32. Lian JB, Stein GS: Concepts of osteoblast growth and differentiation: basis for modulation of bone cell development and tissue formation. *Crit Rev Oral Biol Med*, 3:269-305, 1992.

국문초록

Mineral trioxide aggregate가 유치 및 영구치의 치수기질세포 증식 및 분화에 미치는 영향

김승혜¹ · 전미정¹ · 신동민² · 이제호¹ · 송제선¹연세대학교 치과대학 ¹소아치과학교실, 구강과학연구소, ²구강생물학교실

최근 유구치의 치수절단술 약제로 MTA의 임상 적용이 문헌들에서 보고된 바 있으나 MTA 표면에서 일어나는 유치 치수 세포의 반응에 대한 시험관내 연구는 많이 보고되지 않았다. 이번 연구의 목적은 유치 및 영구치에서 유래한 치수기질세포가 경화된 MTA 표면에서 나타내는 증식 및 분화 능력을 비교 평가하는 것이었다. 사람 영구치와 유치 치수 조직에서 분리된 치수기질세포를 경화된 MTA 표면에서 배양 후 세포증식율과 세포주기를 검사하였으며, 정량적 역전사 중합효소 연쇄반응(RT-PCR)을 사용하여 분화양상을 분석하였다. Runt-related transcription factor 2 (*Runx2*)와 alkaline phosphatase (*ALP*)가 정량적 RT-PCR의 표지자로 사용되었고, MTA 표면에서 증식된 치수기질세포의 형태학적 변화를 주사전자현미경 하에서 관찰하였다. 영구치와 유치의 치수기질세포군은 세포증식률, 세포주기 분포 및 mRNA 발현 양상에 있어서 차이를 보이지 않았으며, 주사전자현미경 상에서 두 군 모두 수지상 형태를 나타내었다. MTA 상에서 관찰된 유치와 영구치의 치수기질세포의 비슷한 증식력 및 광화를 유도하는 세포로의 분화능은 유치의 치수절단술 제재로 MTA가 생체친화적으로 적합함을 보여준다.

주요어: MTA, 치수기질세포, 유치, 영구치, 증식, 분화

www.kci.go.kr